

The Structure of Tautomycin, a Regulator of Eukaryotic Cell Growth

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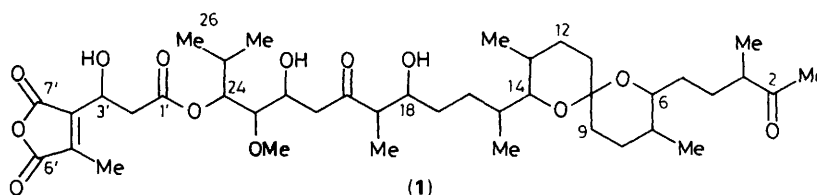
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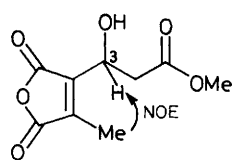
Structure (1) is proposed for tautomycin produced by *Streptomyces spiroverticillatus*, on the basis of chemical degradation and spectroscopic evidences including 2D INADEQUATE of tautomycin labelled with [1,2-¹³C]acetate.

In our laboratory, an antifungal antibiotic, tautomycin, was isolated from a culture of *Streptomyces spiroverticillatus*.¹ Besides antifungal activity, tautomycin induced a morphological change (bleb formation) of human leukaemia cells K562, which is correlated with protein phosphorylation.² On

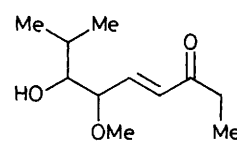
the basis of chemical degradation and spectroscopic evidence, we propose structure (1) for tautomycin.

The field desorption (FD) mass spectrum (m/z 767, MH^+), high resolution positive FAB mass spectrum [m/z 789.4380 ($M + Na$)⁺, Δ 2.1], and the total number of carbons detected by

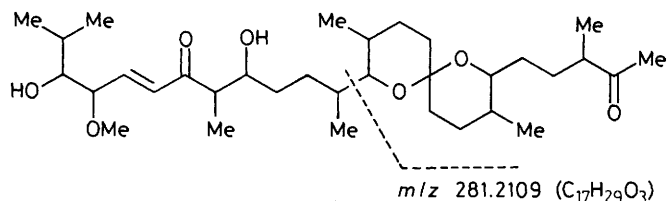




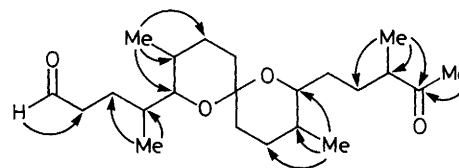
(2a)



(4)



(3)



(5)

^{13}C NMR spectra indicated the molecular formula of $\text{C}_{41}\text{H}_{66}\text{O}_{13}$.[†] UV_{max} (MeCN) at 250 nm and IR bands at 1755 and 1825 cm^{-1} suggested the presence of a 2,3-dialkylmaleic anhydride structure. ^1H NMR of (1) in $[\text{DMSO}-d_6]$ (DMSO = dimethylsulphoxide) showed three hydroxy protons [δ 5.84 (3'-OH), 4.64 (18-OH), 4.46 (22-OH)]. After a week in an NMR tube in $[\text{DMSO}-d_6]$, two carboxyl protons at δ 12.5, and new signals of 3'-H at δ 4.9 and 3'-OH at δ 5.45 appeared. The ratio between the original signals and new signals was approximately 6:4, which remained unchanged even after addition of water.[‡] This data and ^1H - ^1H COSY, ^1H - ^{13}C COSY, and heteronuclear multiple bond correction (HMBC)³ spectra revealed the partial structures (C-18-C-24-C-1'-C-7'), (C-1-C-4), (C-6-C-8), and (C-12-C-15). The quaternary carbon at δ 95.4 suggested the presence of a six membered spiroketal moiety.⁴

Alkaline hydrolysis of (1) (20% Cs_2CO_3 , pH 9, room temp., 2 h) gave an acid (2) and anhydrodeacyltautomycin (3). Treatment of (2) with methanolic H_2SO_4 gave monomethyl ester (2a) and trimethyl ester (2b). Monomethyl ester (2a) [EI mass spectrum (m/z 215, MH^+), IR ($1760, 1830\text{ cm}^{-1}$), UV_{max} (250 nm in MeCN)] showed an equilibrium mixture on HPLC like (1). Difference NOE (3.5% between Me at δ 2.11 and H-3 at δ 4.97, $[\text{DMSO}-d_6]$) and HMBC spectra established the structure of (2a). The IR and UV spectra of (2b) revealed the absence of the 2,3-dialkylmaleic anhydride moiety (disappearance of 1760 and 1830 cm^{-1} in IR and 250 nm UV_{max} in MeCN). No carboxyl proton was detected in the ^1H NMR spectrum of a fresh solution of (2a) in $[\text{DMSO}-d_6]$. After a week in an NMR tube, two carboxyl protons at δ 12.6 appeared and reached an equilibrium.

Both (1) and (2a) exist in methanol-buffer (1% diethylamine-formic acid, pH 7.3) solution as equilibrium mixtures (ca. 5:4). Ring opening and closure of (1) were shown by an oxygen exchange experiment. Treatment of (1) with H_2^{18}O in MeCN, followed by evaporation gave peaks 2, 4, and 6 mass units higher as detected by FD mass spectrometry. The ratio

of incorporation of ^{18}O into (1) increased with time.[§] Similar treatment of (3) with H_2^{18}O gave no higher peaks. Extraction with ethyl acetate at pH 4 resulted in the ring closure.[‡]

The secondary ion (SI) mass spectrum (m/z 567, MH^+) and the total number of carbons detected by ^{13}C NMR spectra of (3) suggested the molecular formula of $\text{C}_{33}\text{H}_{58}\text{O}_7$. High resolution EI mass spectrometry[¶] gave a dehydrated fragment ion [$\text{C}_{33}\text{H}_{54}\text{O}_5$, ($M - 2\text{H}_2\text{O}$)⁺, Δ 1.7 mmu] and an intense fragment ion [m/z 281.2109, ($\text{C}_{17}\text{H}_{29}\text{O}_3$)⁺, Δ 0.5 mmu] as shown. Acetylation of (3) gave diacetate (3a) (m/z 651, MH^+).

Retro-aldol cleavage (20% Cs_2CO_3 , pH 10, room temp., 3 h) of (3) gave (4) and (5). The data from SI MS (m/z 201, MH^+), a homonuclear proton spin decoupling experiment, and ^{13}C NMR established the structure of (4). The coupling constant (17 Hz) between H-4 (δ 6.25, d) and H-5 (δ 6.75, dd) indicated (*E*) configuration of the double bond. The molecular formula ($\text{C}_{22}\text{H}_{38}\text{O}_4$) of (5) was established by high resolution positive FAB MS (m/z 367.2857, MH^+ , Δ 0.9 mmu), negative FAB MS (m/z 365, MH^-)^{||} and the total number of carbons (22C) detected by ^{13}C NMR. HMBC spectra of (5) showed a ^1H - ^{13}C long range coupling pattern as shown.

[§] FD MS; (1) treated with H_2^{18}O for 4 h: ($M + \text{Na}$)⁺, m/z 789 (100%); 791 (80%); 793 (45%); 795 (5%).

[¶] SI MS and high resolution EI MS, Hitachi M80 instrument.

^{||} FAB MS, JMS DX 300 and SX 102 instruments.

^{††} 2D INADEQUATE spectra revealed the correlation of C-1 with C-2, C-2 with C-3, C-3 with 3-Me, C-3 with C-4, C-4 with C-5, C-5 with C-6, C-6 with C-7, C-7 with 7-Me, C-7 with C-8, C-8 with C-9, C-9 with C-10, C-10 with C-11, C-11 with C-12, C-12 with C-13, C-13 with 13-Me, C-13 with C-14, C-14 with C-15, C-15 with 15-Me, C-15 with C-16, C-16 with C-17, C-17 with C-18, C-18 with C-19, C-19 with 19-Me, C-19 with C-20, C-20 with C-21, C-22 with C-23, C-1' with C-2', C-2' with C-3', C-3' with C-4', C-4' with C-7', C-6' with C-5', and C-5' with 5'-Me. Feeding of $[1,2-^{13}\text{C}]$ acetate resulted in considerable randomization in incorporation into (1). Thus, the carbons that should be derived from C-1, C-2, and C-3 of propionate were also enriched. 2D INADEQUATE spectra of $[1,2-^{13}\text{C}]$ acetate labelled tautomycin gave the carbon-carbon connectivity pattern of (1) as described here. A similar randomization was also observed in cationomycin.⁶

[†] FD MS, Hitachi M80 instrument. FAB MS, JMS-SX102. We revised the molecular formula $\text{C}_{42}\text{H}_{70}\text{O}_{12}$ in ref. 1.

[‡] 2,3-Dimethylmaleic anhydride as a model compound also showed similar behaviour. See also ref. 5.

2D INADEQUATE spectroscopy^{††} of tautomycin labelled with [1,2-¹³C]acetate permitted the complete assignments of ¹³C and ¹H NMR signals^{‡‡} and established the total structure. The stereochemistry remains to be solved.

^{‡‡} NMR spectra were measured using JEOL GSX 400 and GSX 500 instruments. All NMR were measured in CDCl₃ except the solvent shown in the text; (**1**), ¹H NMR CDCl₃ [trimethylsilane (TMS)]: 2.12 (s, 3H, H-1), 2.52 (m, 1H, H-3), 1.62 (m, 2H, H-4), 1.25, 1.54 (m, each 2H, H-5), 3.13 (dt, 1H, H-6), 1.24 (m, 1H, H-7), 1.45, 1.58 (m, each 1H, H-8), 1.45, 1.62 (m, each 1H, H-9), 1.38, 1.55 (m, each 1H, H-11), 1.38, 2.00 (m, each 1H, H-12), 1.82 (m, 1H, H-13), 3.25 (dd, 1H, H-14), 1.54 (m, 1H, H-15), 1.52, 1.83 (m, each 1H, H-16), 1.27, 1.61 (m, each 1H, H-17), 3.68 (m, 1H, H-18), 2.62 (m, 1H, H-19), 2.63, 2.97 (m, each 1H, H-21), 4.33 (ddd, 1H, H-23), 3.25 (dd, 1H, H-23), 5.07 (dd, 1H, H-24), 2.08 (m, 1H, H-25), 0.95 (d, 3H, H-26), 1.07 (d, 3H, 3-Me), 0.8 (d, 3H, 7-Me), 0.9 (d, 3H, 13-Me), 0.95 (d, 3H, 15-Me), 1.08 (d, 3H, 19-Me), 0.95 (d, 3H, 25-Me), 3.42 (s, 3H, 23-OMe), 2.75, 2.90 (dd, each 1H, H-2'), 5.18 (dd, 1H, H-3'), 2.24 (s, 3H, 5'-Me).

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